

IL-33/ST2 Signaling Regulates Synaptic Plasticity and Homeostasis in the Adult Hippocampal Circuitry

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In response to neuronal activity changes, the adult hippocampal circuits undergo continuous synaptic remodeling, which is essential for information processing, learning, and memory encoding. Glial cells, including astrocytes and microglia, actively regulate hippocampal synaptic plasticity by coordinating the neuronal activity-induced synaptic changes at the circuit level. Emerging evidence suggests that the crosstalk between neurons and glia in the adult hippocampus is region specific and that the mechanisms controlling this process are critically dependent on secreted factors. Interleukin-33 (IL-33), a cytokine of the IL-1 family, is a key factor that modulates such glia-driven neuromodulations in two distinct hippocampal circuits. The activation of IL-33 and its receptor complex is important for maintaining the excitatory synaptic activity in the cornu ammonis 1 subregion and the remodeling of dentate gyrus synapses through activity-dependent astrocyte–synapse and microglia–synapse interactions, respectively. Meanwhile, the dysregulation of this signaling is implicated in multiple neurological disorders, especially Alzheimer’s disease. Further investigations of how IL-33/ST2 signaling is regulated in a region-specific manner as well as its diverse functions in glia–synapse communications in the adult hippocampal circuitry will provide insights into the nature of hippocampal synaptic plasticity and homeostasis in health and disease.

Keywords: interleukin, astrocyte, microglia, neuronal activity, learning and memory, Alzheimer’s disease

Introduction

THE HIPPOCAMPUS IS critical for information encoding as well as memory storage and retrieval. It can be divided into the cornu ammonis 1 (CA1), CA2, CA3, and dentate gyrus (DG) subregions (Basu and Siegelbaum, 2015). Of note, the DG–CA3–CA1 synaptic circuit is essential for hippocampus-dependent memory formation in response to experience (Basu and Siegelbaum, 2015). The neurons in this circuit can alter the structure and strength of synapses in a neuronal activity-dependent manner, which is termed “synaptic plasticity”. Such activity-driven regulation of synaptic efficacy ultimately shapes the connectivity of neuronal circuits in the adult hippocampus, thereby facilitating learning and memory (Bannerman *et al.*, 2014; Magee and Grienberger, 2020). In particular, homeostatic plasticity is a unique form of synaptic plasticity, wherein neurons adapt to offset excessive excitation or inhibition induced by chronic changes in synaptic activity, which is important for maintaining circuit homeostasis (Turrigiano, 2008; Pozo and Goda, 2010). However, improper induction and expression

of hippocampal synaptic plasticity contribute to the synaptic dysfunction and memory deficits in several neurodegenerative diseases, including Alzheimer’s disease (AD) (Chen *et al.*, 2019). Thus, to better understand how the hippocampus handles information as well as its related memory functions in both physiological and pathological conditions, detailed knowledge about the synaptic plasticity in distinct hippocampal circuits is required.

Emerging evidence suggests that hippocampal synaptic plasticity is regulated not only by the bidirectional communication between presynaptic and postsynaptic neurons but also through the interactions between neurons and their enveloping glia (Allen and Lyons, 2018; Wang *et al.*, 2021a). Astrocytes and microglia, the major types of glial cells in the adult hippocampus, can interact with synapses through their intricate processes (Perea *et al.*, 2009; Perez-Alvarez *et al.*, 2014; Allen and Lyons, 2018). In particular, the specialized processes of different glia express distinct receptors, which enable them to detect changes in synaptic activity in a specific circuit, thereby enabling these cells to regulate hippocampal synaptic plasticity in a subregion-specific manner

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(Oliet *et al.*, 2001; Bernardinelli *et al.*, 2014; De Pitta *et al.*, 2016). Therefore, activation of proper glial signaling is critical for the circuit homeostasis and functioning of adult hippocampus, whereas the dysregulation of glial signaling is implicated in multiple neurodegenerative diseases (Phatnani and Maniatis, 2015; von Bernhardt *et al.*, 2015; Wang *et al.*, 2021a). However, it remains unclear how glia sense changes in neuronal activity and consequently affect hippocampal circuit connectivity. Nevertheless, it was recently proposed that the cytokine interleukin-33 (IL-33) is involved in such glia–neuron communication in the adult hippocampal circuitry and that IL-33-mediated signals are important for hippocampal synaptic plasticity and homeostasis (Nguyen *et al.*, 2020; Wang *et al.*, 2021b).

IL-33 was initially identified as an alarmin of the IL-1 family crucial for maintaining tissue homeostasis by regulating both innate and adaptive immune responses (Liew *et al.*, 2010, 2016). Upon release, IL-33 binds to its receptor complex comprising ST2 and IL-1RAcP (IL-1 receptor accessory protein) to activate the nuclear factor-kappa B and mitogen-activated protein kinase pathways, thereby driving multiple biological processes (Liew *et al.*, 2016). Besides the basic functions of IL-33 in immune responses, recent findings have extended the physiological roles of IL-33 to include synaptic remodeling. Specifically, IL-33 exhibits enriched expression in the neural cells in the adult hippocampus and regulates hippocampal synaptic functions in an activity- and circuit-dependent manner (Nguyen *et al.*, 2020; Wang *et al.*, 2021b).

Astrocyte-Secreted IL-33 Regulates Synaptic Homeostasis in the Hippocampal CA3–CA1 Circuit in an Activity-Dependent Manner

We recently identified an activity-dependent, IL-33-mediated interaction between astrocytes and neurons in the adult hippocampal CA3–CA1 circuit (Fig. 1A, B) (Wang *et al.*, 2021b). Pharmacological blockade of neuronal activity by tetrodotoxin increased IL-33 expression and secretion in acute hippocampal slices. Of note, such activity-dependent regulation of IL-33 only occurred in a specific subpopulation of hippocampal CA1 astrocytes, suggesting that astrocyte-secreted IL-33 might mediate the activity blockade-induced hippocampal homeostatic plasticity in a circuit-specific manner. Moreover, upon optogenetic stimulation to inactivate hippocampal CA1 excitatory neurons *in vivo* by directly suppressing their activity or activating their neighboring parvalbumin interneurons (Klausberger and Somogyi, 2008; Ferguson and Gao, 2018), we still observed significantly increased IL-33 expression in CA1 astrocytes, suggesting that astrocyte-derived IL-33 has a unique role in the regulation of synaptic connectivity and homeostasis in the adult hippocampal CA1 circuitry.

Indeed, IL-33 administration in young adult mice promotes excitatory synapse formation and enhances synaptic transmission in the CA1 excitatory neurons. Accordingly, this synaptogenic effect of IL-33 is mediated by ST2-mediated postsynaptic mechanisms, including the phosphorylation-dependent synaptic accumulation of the postsynaptic scaffold protein PSD-95 and the subsequent strengthening of synaptic transmission. Given that chronic changes in the abundance of postsynaptic PSD-95 regulated by Ser295

phosphorylation possibly contribute to homeostatic plasticity (Kim *et al.*, 2007), activity-induced astrocyte-secreted IL-33 might be a critical signaling regulator of this process. Meanwhile, conditional knockout of IL-33, specifically in CA1 astrocytes, decreases the number of excitatory synapses, further indicating that astrocyte-secreted IL-33 is required for the synaptic remodeling of the hippocampal CA3–CA1 circuit. These findings reveal a cellular process, whereby hippocampal CA1 astrocytes can sense and respond to cues from the CA3–CA1 circuit by the activation of IL-33 signaling. Nonetheless, it remains unclear how IL-33 expression or secretion is regulated in the astrocytes surrounding CA1 excitatory synapses or how it exerts synaptogenic effects during learning and memory. Given that diverse astrocytic ion channels and neurotransmitter receptors are responsible for the activity-dependent astrocyte–synapse interactions (Olsen *et al.*, 2015; Allen and Lyons, 2018), further efforts to determine the specific receptors involved in this process would help explain this activity-driven IL-33 regulation in CA1 astrocytes.

Our study revealed an IL-33-driven synaptic adaptation event in the hippocampal CA1 circuit wherein activity deprivation of CA1 excitatory neurons stimulates IL-33 secretion from neighboring astrocytes; this secreted IL-33 acts on its neuronal receptor complex to regulate the homeostatic synaptic plasticity in these excitatory neurons by increasing the numbers of structural and functional excitatory synapses, thereby maintaining hippocampal network stability (Fig. 1B). Besides IL-33, studies including our RNA sequencing analysis have identified several secreted factors that are regulated by neuronal activity and involved in synaptic functions, including Chrdl1, SPARCL1, and TNF α (Stellwagen and Malenka, 2006; Kucukdereli *et al.*, 2011; Blanco-Suarez *et al.*, 2018; Wang *et al.*, 2021b). Thus, IL-33 might coordinate with multiple well-known cellular signals to constitute a homeostatic feedback loop whereby CA1 neurons increase synaptic excitability, enabling them to receive more inputs from CA3 presynaptic neurons during activity inhibition; this would not only help maintain CA3–CA1 synaptic homeostasis but also facilitate a stable circuit activity in preparation for the subsequent encoding of learning experiences. As such, further examination of the activity-induced signaling cascades between astrocytes and neurons in the CA1 hippocampal circuitry will help clarify the nature of hippocampal homeostatic plasticity. Concordantly, perturbation of this IL-33-dependent homeostatic regulation by the IL-33 decoy receptor, soluble ST2, results in hippocampus-associated cognitive impairments (Wang *et al.*, 2021b). Hence, astrocyte-secreted IL-33 is a key synaptic regulator of hippocampal homeostatic plasticity, and the homeostasis of the CA3–CA1 circuit maintained by such astrocyte–neuron IL-33/ST2 signaling is required for information processing in the adult hippocampus.

Neuron–Microglia IL-33/ST2 Signaling Is Required for Experience-Dependent Synaptic Remodeling of the Hippocampal DG Circuit

In addition to our finding that astrocyte-secreted IL-33 exhibits a region-specific effect on hippocampal homeostatic plasticity, Nguyen *et al.* demonstrated that in response to experience (i.e., an enriched environment), IL-33 mRNA

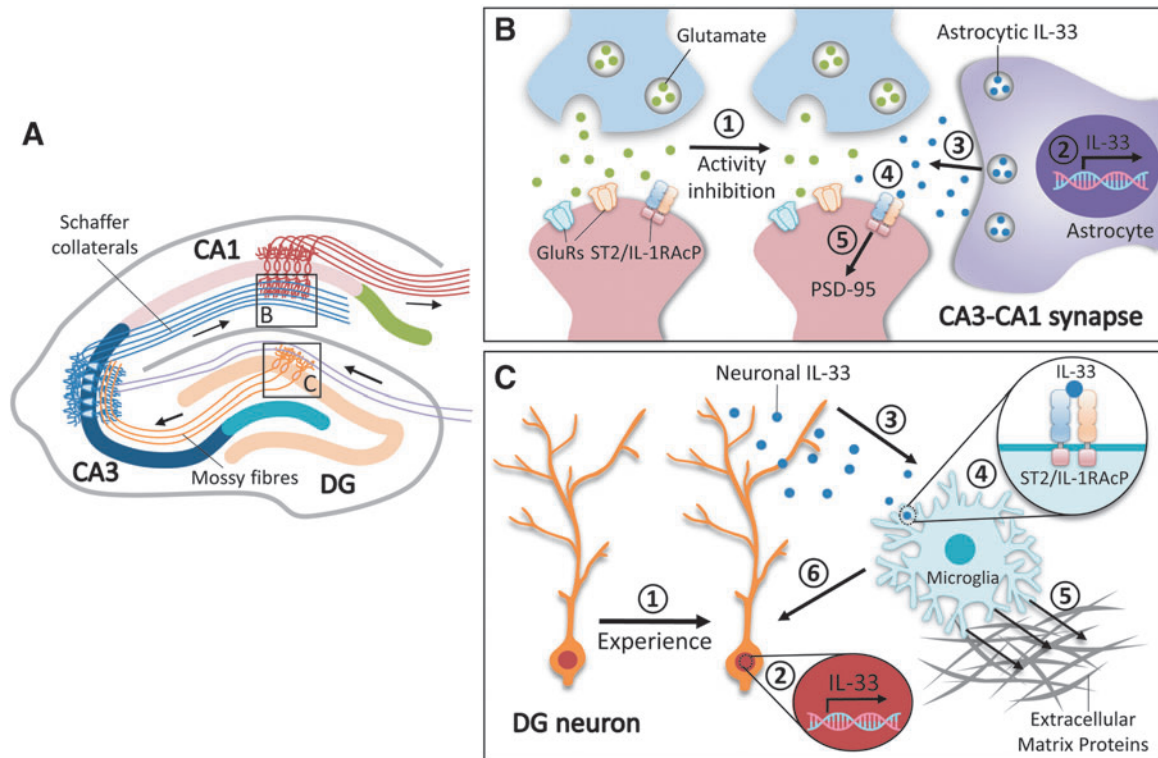


FIG. 1. IL-33/ST2 signaling regulates hippocampal synaptic plasticity and homeostasis through circuit-dependent gliasynapse interactions. **(A)** Schematic representation of the hippocampal DG–CA3–CA1 synaptic circuit. During sensory experience, DG granule neurons receive synaptic inputs from the entorhinal cortex and then send their projections to CA3 pyramidal neurons through mossy fibers, which subsequently project to CA1 pyramidal neurons through the Schaffer collaterals. This constitutes a pathway whereby sensory information is integrated in CA1 pyramidal neurons and eventually leaves the hippocampus through the CA1 subregion to other brain regions. **(B)** In the adult hippocampal CA3–CA1 circuit, astrocyte-secreted IL-33 is required for activity blockade-induced homeostatic plasticity. Upon inhibition of CA3–CA1 excitatory synaptic activity (1), IL-33 expression is increased in astrocytes surrounding synapses (2) and IL-33 is then secreted (3) and binds to the postsynaptic ST2/IL-1RAcP receptor complex (4) to enhance the synaptic accumulation of the postsynaptic scaffold protein PSD-95 and the subsequent strengthening of synaptic transmission (5). Such activity-driven feedback control of CA3–CA1 synaptic strength is important for maintaining the functioning and homeostasis of the adult hippocampal circuitry. **(C)** In the adult hippocampal DG circuit, neuronal IL-33 regulates experience-driven synaptic remodeling through microglial phagocytosis. In response to experience (1), DG neurons exhibit increased expression (2) and secretion (3) of IL-33, which in turn binds to the microglial ST2/IL-1RAcP receptor complex (4) to induce the phagocytosis of extracellular matrix proteins (5), thereby promoting synaptic remodeling and plasticity in the DG circuit (6). CA, cornu ammonis; DG, dentate gyrus; GluRs, glutamate receptors; IL, interleukin.

expression is increased in a subtype of hippocampal neurons in the DG circuit; in turn, IL-33 protein binds to its ST2 receptor in nearby microglia to induce the phagocytosis of extracellular matrix proteins (Fig. 1A, C) (Nguyen *et al.*, 2020). Given that extracellular matrix proteins can restrict the induction and expression of synaptic plasticity (Frischknecht *et al.*, 2009), such clearance of extracellular matrix proteins during experience due to neuron–microglia IL-33/ST2 signaling is likely essential for synapse formation and plasticity in the DG, demonstrating the importance of such signaling for memory performance in young adult mice. Accordingly, dysregulated neuron–microglia IL-33/ST2 signaling in the adult hippocampal DG is implicated in aging, which is associated with memory deficits (Nguyen *et al.*, 2020). Meanwhile, the activation of IL-33/ST2 signaling in the aging mouse brain rescues synaptic dysfunctions. Nevertheless, it remains unclear how DG neurons transduce external experience into the internal signaling cascade that includes the secretion of neuronal IL-33 and the

subsequent activation of adjacent microglial phagocytosis. Given that microglia-mediated engulfment and clearance of excessive synapses (i.e., synaptic pruning) are critical for circuit development and maintenance (Paolicelli *et al.*, 2011), it is of interest to further examine why such enriched environment-induced microglial phagocytosis only targets matrix proteins and not synapses. In addition, the DG–CA3–CA1 synaptic circuit is critical for information encoding during learning experience (Basu and Siegelbaum, 2015). Thus, determining the effects induced by enriched environments in other hippocampal subregions, including the CA3 and CA1, would help clarify the nature of such activity-dependent remodeling of hippocampal circuits.

Region-Specific Control of Adult Hippocampal Circuit Homeostasis by IL-33/ST2 Signaling

In the adult brain, IL-33 is abundantly expressed by astrocytes and oligodendrocytes (Hudson *et al.*, 2008; Yasuoka

et al., 2011). The expression patterns and levels of IL-33 differ among brain regions owing to their distinct transcriptional mechanisms (Fairlie-Clarke *et al.*, 2018), including the expression of different enriched transcription factors (Lozzi *et al.*, 2020) and dynamic chromatin modulations (Klemm *et al.*, 2019). Such differential expression of IL-33 suggests that it might have distinct functions in neuronal circuits in certain hippocampal regions. Indeed, we found that the IL-33 secreted by a subpopulation of astrocytes in the hippocampal CA1 region is specifically involved in activity blockade-induced homeostatic synaptic plasticity (Fig. 1B) (Wang *et al.*, 2021b). Meanwhile, IL-33 secreted by neurons in the DG regulates synaptic remodeling by microglial engulfment (Fig. 1C) (Nguyen *et al.*, 2020). Thus, these findings collectively reveal a key role of IL-33 in the activity- and circuit-specific control of synaptic functions in the adult hippocampus.

Moreover, the recent emergence of novel single-cell techniques has revealed that neural cells, especially astrocytes and microglia, have highly heterogeneous genetic profiles, morphology, and functions—even within the same brain region (Mederos *et al.*, 2018; Masuda *et al.*, 2020). The heterogeneity of neural cells might explain their different responses to the activity-induced expression and secretion of IL-33, namely the postsynaptic responses in the CA1 subregion that regulate homeostatic plasticity as well as the microglial activation in the DG that modulates synaptic remodeling. Such distinct mechanisms elicited by IL-33/ST2 signaling enable diverse avenues for the homeostatic regulation of synaptic activity and functions in the adult hippocampal circuits in response to different learning experiences. Therefore, further single-cell transcriptomic profiling of different neural cell types in various hippocampal subregions would help reveal the cellular basis of the activity-driven neuron–glia communication in the adult hippocampal circuitry, thereby providing insights into the molecular mechanisms by which glia modulate hippocampal synaptic plasticity.

Dysregulation of IL-33/ST2 Signaling in AD

Of note, both of the abovementioned studies on IL-33-mediated glia–synapse interactions imply that dysregulated hippocampal IL-33/ST2 signaling is associated with memory deficits (Nguyen *et al.*, 2020; Wang *et al.*, 2021b). Concordantly, IL-33/ST2 signaling dysfunction is implicated in the progression of several neurological disorders such as AD (Chapuis *et al.*, 2009; Allan *et al.*, 2016; Fu *et al.*, 2016; Liew *et al.*, 2016). Accordingly, human genetics studies have revealed several IL-33 single-nucleotide polymorphisms related to AD (Chapuis *et al.*, 2009). Compared to healthy controls, patients with mild cognitive impairment or AD have fewer synapses in the hippocampus (Dickerson *et al.*, 2005; Yassa *et al.*, 2010) and decreased IL-33/ST2 signaling in the central nervous system (Chapuis *et al.*, 2009; Fu *et al.*, 2016). Given that both hyperactivated microglia and astrocytes have been observed in the brains of patients with AD and transgenic mouse models of AD (Keren-Shaul *et al.*, 2017; Fakhoury, 2018; Hemonnot *et al.*, 2019; Habib *et al.*, 2020), the dysregulated IL-33 signaling observed in AD is likely involved in the impaired glia–synapse interactions in the adult hippocampal circuitry.

Therefore, it is of great interest to further investigate the functional roles of IL-33 signaling in glia-driven synaptic remodeling in AD. Of note, we found that replenishment of IL-33 in a transgenic mouse model of amyloid deposition (i.e., APP/PS1 mice) ameliorates synaptic plasticity deficits and cognitive impairment (Fu *et al.*, 2016), which suggests that IL-33 indeed has a beneficial effect against AD pathology.

Conclusion and Perspectives

The findings discussed above collectively suggest the existence of a diverse IL-33/ST2 signaling-mediated biological event in the brain that extends its functions to immune cells, thereby regulating glia–neuron communication and impacting synaptic plasticity and homeostasis in the adult hippocampal circuitry. Thus, the coordination of distinct functions of IL-33/ST2 signaling in different hippocampal subregions is essential for experience-driven information processing in the adult hippocampus. Accordingly, further investigation of the molecular and cellular mechanisms of the IL-33/ST2 signaling-driven synaptic adaptations in neuronal circuits will advance our understanding of hippocampus-associated memory processes, which will also provide insights into the development of therapeutic strategies for cognitive deficits in neurological disorders.

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No competing financial interests exist.

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References

- Allan, D., Fairlie-Clarke, K.J., Elliott, C., Schuh, C., Barnett, S.C., Lassmann, H., *et al.* (2016). Role of IL-33 and ST2 signalling pathway in multiple sclerosis: expression by oligodendrocytes and inhibition of myelination in central nervous system. *Acta Neuropathol Commun* **4**, 75.
- Allen, N.J., and Lyons, D.A. (2018). Glia as architects of central nervous system formation and function. *Science* **362**, 181–185.
- Bannerman, D.M., Sprengel, R., Sanderson, D.J., McHugh, S.B., Rawlins, J.N., Monyer, H., *et al.* (2014). Hippocampal synaptic plasticity, spatial memory and anxiety. *Nat Rev Neurosci* **15**, 181–192.

- Basu, J., and Siegelbaum, S.A. (2015). The corticohippocampal circuit, synaptic plasticity, and memory. *Cold Spring Harb Perspect Biol* **7**, a021733.
- Bernardinelli, Y., Randall, J., Janett, E., Nikonenko, I., König, S., Jones, E.V., *et al.* (2014). Activity-dependent structural plasticity of perisynaptic astrocytic domains promotes excitatory synapse stability. *Curr Biol* **24**, 1679–1688.
- Blanco-Suarez, E., Liu, T.F., Kopelevich, A., and Allen, N.J. (2018). Astrocyte-secreted chordin-like 1 drives synapse maturation and limits plasticity by increasing synaptic GluA2 AMPA receptors. *Neuron* **100**, 1116–1132.e13.
- Chapuis, J., Hot, D., Hansmann, F., Kerdraon, O., Ferreira, S., Hubans, C., *et al.* (2009). Transcriptomic and genetic studies identify IL-33 as a candidate gene for Alzheimer's disease. *Mol Psychiatr* **14**, 1004–1016.
- Chen, Y., Fu, A.K.Y., and Ip, N.Y. (2019). Synaptic dysfunction in Alzheimer's disease: mechanisms and therapeutic strategies. *Pharmacol Ther* **195**, 186–198.
- De Pitta, M., Brunel, N., and Volterra, A. (2016). Astrocytes: orchestrating synaptic plasticity? *Neuroscience* **323**, 43–61.
- Dickerson, B.C., Salat, D.H., Greve, D.N., Chua, E.F., Rand-Giovannetti, E., Rentz, D.M., *et al.* (2005). Increased hippocampal activation in mild cognitive impairment compared to normal aging and AD. *Neurology* **65**, 404–411.
- Fairlie-Clarke, K., Barbour, M., Wilson, C., Hridi, S.U., Allan, D., and Jiang, H.R. (2018). Expression and function of IL-33/ST2 axis in the central nervous system under normal and diseased conditions. *Front Immunol* **9**, 2596.
- Fakhoury, M. (2018). Microglia and astrocytes in Alzheimer's disease: implications for therapy. *Curr Neuropharmacol* **16**, 508–518.
- Ferguson, B.R., and Gao, W.J. (2018). PV interneurons: critical regulators of E/I balance for prefrontal cortex-dependent behavior and psychiatric disorders. *Front Neural Circuits* **12**, 37.
- Frischknecht, R., Heine, M., Perrais, D., Seidenbecher, C.I., Choquet, D., and Gundelfinger, E.D. (2009). Brain extracellular matrix affects AMPA receptor lateral mobility and short-term synaptic plasticity. *Nat Neurosci* **12**, 897–904.
- Fu, A.K., Hung, K.W., Yuen, M.Y., Zhou, X., Mak, D.S., Chan, I.C., *et al.* (2016). IL-33 ameliorates Alzheimer's disease-like pathology and cognitive decline. *Proc Natl Acad Sci U S A* **113**, E2705–E2713.
- Habib, N., McCabe, C., Medina, S., Varshavsky, M., Kitsberg, D., Dvir-Szternfeld, R., *et al.* (2020). Disease-associated astrocytes in Alzheimer's disease and aging. *Nat Neurosci* **23**, 701–706.
- Hemonnot, A.L., Hua, J., Ulmann, L., and Hirbec, H. (2019). Microglia in Alzheimer disease: well-known targets and new opportunities. *Front Aging Neurosci* **11**, 233.
- Hudson, C.A., Christophi, G.P., Gruber, R.C., Wilmore, J.R., Lawrence, D.A., and Massa, P.T. (2008). Induction of IL-33 expression and activity in central nervous system glia. *J Leukoc Biol* **84**, 631–643.
- Keren-Shaul, H., Spinrad, A., Weiner, A., Matcovitch-Natan, O., Dvir-Szternfeld, R., Ulland, T.K., *et al.* (2017). A unique microglia type associated with restricting development of Alzheimer's disease. *Cell* **169**, 1276–1290.e1217.
- Kim, M.J., Futai, K., Jo, J., Hayashi, Y., Cho, K., and Sheng, M. (2007). Synaptic accumulation of PSD-95 and synaptic function regulated by phosphorylation of serine-295 of PSD-95. *Neuron* **56**, 488–502.
- Klausberger, T., and Somogyi, P. (2008). Neuronal diversity and temporal dynamics: the unity of hippocampal circuit operations. *Science* **321**, 53–57.
- Klemm, S.L., Shipony, Z., and Greenleaf, W.J. (2019). Chromatin accessibility and the regulatory epigenome. *Nat Rev Genet* **20**, 207–220.
- Kucukdereli, H., Allen, N.J., Lee, A.T., Feng, A., Ozlu, M.I., Conatser, L.M., *et al.* (2011). Control of excitatory CNS synaptogenesis by astrocyte-secreted proteins Hevin and SPARC. *Proc Natl Acad Sci U S A* **108**, E440–E449.
- Liew, F.Y., Girard, J.P., and Turnquist, H.R. (2016). Interleukin-33 in health and disease. *Nat Rev Immunol* **16**, 676–689.
- Liew, F.Y., Pitman, N.I., and McInnes, I.B. (2010). Disease-associated functions of IL-33: the new kid in the IL-1 family. *Nat Rev Immunol* **10**, 103–110.
- Lozzi, B., Huang, T.W., Sardar, D., Huang, A.Y., and Deneen, B. (2020). Regionally distinct astrocytes display unique transcription factor profiles in the adult brain. *Front Neurosci* **14**, 61.
- Magee, J.C., and Grienberger, C. (2020). Synaptic plasticity forms and functions. *Annu Rev Neurosci* **33**, 18–41.
- Masuda, T., Sankowski, R., Staszewski, O., and Prinz, M. (2020). Microglia Heterogeneity in the Single-Cell Era. *Cell Rep* **30**, 1271–1281.
- Mederos, S., Gonzalez-Arias, C., and Perea, G. (2018). Astrocyte-neuron networks: a multilane highway of signaling for homeostatic brain function. *Front Synaptic Neurosci* **10**, 45.
- Nguyen, P.T., Dorman, L.C., Pan, S., Vainchtein, I.D., Han, R.T., Nakao-Inoue, H., *et al.* (2020). Microglial remodeling of the extracellular matrix promotes synapse plasticity. *Cell* **182**, 388–403.e15.
- Oliet, S.H.R., Piet, R., and Poulain, D.A. (2001). Control of glutamate clearance and synaptic efficacy by glial coverage of neurons. *Science* **292**, 923–926.
- Olsen, M.L., Khakh, B.S., Skatchkov, S.N., Zhou, M., Lee, C.J., and Rouach, N. (2015). New insights on astrocyte ion channels: critical for homeostasis and neuron-glia signaling. *J Neurosci* **35**, 13827–13835.
- Paolicelli, R.C., Bolasco, G., Pagani, F., Maggi, L., Scianni, M., Panzanelli, P., *et al.* (2011). Synaptic pruning by microglia is necessary for normal brain development. *Science* **333**, 1456–1458.
- Perea, G., Navarrete, M., and Araque, A. (2009). Tripartite synapses: astrocytes process and control synaptic information. *Trends Neurosci* **32**, 421–431.
- Perez-Alvarez, A., Navarrete, M., Covelo, A., Martin, E.D., and Araque, A. (2014). Structural and functional plasticity of astrocyte processes and dendritic spine interactions. *J Neurosci* **34**, 12738–12744.
- Phatnani, H., and Maniatis, T. (2015). Astrocytes in neurodegenerative disease. *Cold Spring Harb Perspect Biol* **7**, a020628.
- Pozo, K., and Goda, Y. (2010). Unraveling mechanisms of homeostatic synaptic plasticity. *Neuron* **66**, 337–351.
- Stellwagen, D., and Malenka, R.C. (2006). Synaptic scaling mediated by glial TNF- α . *Nature* **440**, 1054–1059.
- Turrigiano, G.G. (2008). The self-tuning neuron: synaptic scaling of excitatory synapses. *Cell* **135**, 422–435.
- von Bernhardi, R., Eugenin-von Bernhardi, L., and Eugenin, J. (2015). Microglial cell dysregulation in brain aging and neurodegeneration. *Front Aging Neurosci* **7**, 124.
- Wang, Y., Fu, A.K.Y., and Ip, N.Y. (2021a). Instructive roles of astrocytes in hippocampal synaptic plasticity: neuronal activity-dependent regulatory mechanisms. *FEBS J* [Epub ahead of print]; DOI: 10.1111/febs.15878.

- Wang, Y., Fu, W.Y., Cheung, K., Hung, K.W., Chen, C., Geng, H., *et al.* (2021b). Astrocyte-secreted IL-33 mediates homeostatic synaptic plasticity in the adult hippocampus. *Proc Natl Acad Sci U S A* **118**, e2020810118.
- Yassa, M.A., Stark, S.M., Bakker, A., Albert, M.S., Gallagher, M., and Stark, C.E. (2010). High-resolution structural and functional MRI of hippocampal CA3 and dentate gyrus in patients with amnesic Mild Cognitive Impairment. *Neuroimage* **51**, 1242–1252.
- Yasuoka, S., Kawanokuchi, J., Parajuli, B., Jin, S., Doi, Y., Noda, M., *et al.* (2011). Production and functions of IL-33 in the central nervous system. *Brain Res* **1385**, 8–17.

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